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BRIEF ARTICLE

α -fetoprotein, vascular endothelial growth factor receptor-1 and early recurrence of hepatoma

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Abstract

AIM: To investigate whether α -fetoprotein (AFP) and vascular endothelial growth factor receptor (VEGFR)-1 correlate with early recurrence of hepatoma/hepatocellular carcinoma (HCC).

METHODS: From 2000 to 2005, 114 consecutive patients with HCC underwent primary curative hepatectomy. The mean age was 60.7 (8.7) years and 94 patients were male. The median follow-up period was 71.2 mo (range: 43-100 mo). Immediately prior to commencing laparotomy, 5 mL bone marrow was aspirated from the

sternum and collected in citrate-coated test tubes. The initial 2 mL of bone marrow aspirate was discarded in each case. AFP mRNA and VEGFR-1 mRNA in the bone marrow and peripheral blood (BM- and PH-AFP mRNA and BM- and PH-VEGFR-1 mRNA, respectively) were measured by real-time quantitative reverse transcription polymerase chain reaction. As normal controls, VEGFR-1 mRNA in the bone marrow and peripheral blood was also measured in 11 living liver donors. These data were evaluated for any correlation with early recurrence, comparing clinical and pathological outcomes.

RESULTS: The cut-off value of the BM-AFP mRNA and PH-AFP mRNA level in patients with HCC was set at 1.92 \times 10⁻⁷ and zero, respectively, based on data from the controls. A total of 34 (29.8%) and six (5.4%) patients were positive for BM-AFP mRNA and PH-AFP mRNA, respectively. The BM-VEGFR-1 mRNA levels in all HCC patients were higher than those in the normal controls, and this was the case also for PH-VEGFR-1mRNA. The 25-percentile values for the BM- and PH-VEGFR-1 mRNA in HCC patients were used as the cut-off values for assigning the patients into two groups based on these transcript levels. The High group for BM- VEG-FR-1 mRNA contained 81 (71.1%) HCC cases and the Low group was assigned 33 (28.9%) patients. These numbers for PH-VEGFR-1mRNA were 78 (75.0%) and 26 (25.0%), respectively. HCC recurred in 80 patients; in the remnant liver in 48 cases, in the remnant liver and remote tissue in 20, and in the remote tissue alone in 12. BM-AFP mRNA-positive cases showed a significantly higher rate of early recurrence (within 1 year of surgical treatment) compared with BM-AFP mRNAnegative patients (P = 0.0091). Patients were classified into four groups according to the level/status of their BM-VEGFR-1 and BM-AFP mRNA as follows: group A (n = 23), BM-VEGFR-1/BM-AFP mRNA = low/negative; group B (n = 57) high/negative; group C (n = 10) low/ positive; group D (n = 24), high/positive. This classification was found to correlate with a recurrence of this



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disease within 1 year (P = 0.0228). The disease-free survival curve of group A was significantly better than that of groups B, C or D (P = 0.0437, P = 0.0325, P = 0.0225). No other classification (i.e., PH-VEGF-R1/BM-AFP, BM-VEGF-R1/PH-AFP, and PH-VEGF-R1/PH-AFP mRNA) showed such a correlation.

CONCLUSION: The evaluation of BM-AFP and BM-VEG-FR-1 mRNA in patients with HCC may be a valuable predictor of disease recurrence following curative resection.

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Key words: α -fetoprotein; Vascular endothelial growth factor receptor-1; mRNA; Early recurrence; Hepatocellular carcinoma

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INTRODUCTION

Various factors are thought to contribute to hepatocellular carcinoma (HCC) recurrence, which commonly results in death, including multicentric carcinogenesis in the remnant liver due to an underlying hepatitis-B-virus- or hepatitis-C-virus-induced liver cirrhosis^[1], hematogenic spread, or micrometastasis of HCC cells prior to surgery or during hepatectomy by manipulation of the liver^[2]. Recently, using various molecular biological markers, the detection of malignant cells in the systemic circulation and bone marrow has become possible and the presence of these cells has been found to correlate with the clinical outcome^[3-8]. We have also reported from our laboratory that the detection of HCC cells in the bone marrow by real-time reverse-transcriptase polymerase chain reaction (RT-PCR) analysis of α -fetoprotein (AFP) mRNA before curative hepatectomy correlates with HCC recurrence and patient survival outcomes. Although early recurrence within 1 year of curative resection for HCC is one of the most important factors affecting the prognosis and clinical outcomes^[9,10], the relationship between early recurrence and disseminated cancer cells has not yet been evaluated.

It has been recently hypothesized that metastasis is dependent on both isolated cancer cells and the host response. Kaplan *et al*¹¹¹ have reported that bone-marrowderived hematopoietic progenitor cells that express vascular endothelial growth factor receptor (VEGFR)-1 migrate to tumor-specific pre-metastatic sites and form cellular clusters before the arrival of tumor cells both *in vitro* and *in vivo*. Moreover, it has been reported that the simultaneous presence of isolated tumor cells and VEGFR-1 expression at pre-metastatic sites is clinically significant for disease progression in gastric cancer^[12]. With regard to HCC however, there has been no study to date of the association between isolated cancer cells and the expression of VEGFR-1.

In our present study, we examined whether the expression of AFP mRNA and VEGFR-1 in the bone marrow and peripheral blood, detected by sensitive real-time quantitative RT-PCR, could predict early recurrence in consecutive HCC patients who had undergone a curative hepatic resection.

MATERIALS AND METHODS

Ethics

This study was approved by the Institutional Review Board of the Hokkaido University, School of Advanced Medicine. Informed consent was obtained from each patient in accordance with the Ethics Committee Guidelines at our institution.

From July 2000 to June 2005, 114 consecutive patients underwent primary curative hepatectomy at the First Department of Surgery, Hokkaido University Hospital. The mean age was 60.7 (8.7) years and 94 patients were male. The Child-Pugh staging was A in 110 patients and B in four. Patients were discharged from the hospital at an average of 17.5 (7.1) d after surgery. They were followed up at 3-mo intervals by computed tomography (CT), magnetic resonance imaging (MRI), ultrasonography (US) and laboratory tests for AFP, lens culinaris agglutinin-reactive fraction of AFP (AFP-L3), and protein induced by vitamin K absence or antagonist-II (PIVKA-II). The median follow-up period was 71.2 mo (range: 43 mo-100 mo).

As normal controls, VEGFR-1 mRNA in the bone marrow and peripheral blood was also measured in 11 living liver donors. The cut-off value for AFP mRNA/glyceraldehyde-3-phosphate dehydrogenase (GAPDH) in the bone marrow and peripheral blood was set as described in our previous study^[13].

Sample collections

Immediately prior to commencing the laparotomy, 5 mL bone marrow was aspirated from the sternum and collected in citrate-coated test tubes. The initial 2 mL of the bone marrow aspirate was discarded in each case.

RNA isolation and reverse transcription

Bone marrow samples were prepared for the measurement of total RNA using a Blood RNA extraction kit (QIAGEN, Hilden, Germany) according to the manufacturer's protocol with minor modifications. Briefly, 5 mL bone marrow cells were mixed with 25 mL Reagent buffer erythrocyte lysis (EL). They were then cooled on

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ice for 15 min, centrifuged, and the cell pellets were collected. The pellets were suspended in 1.35 mL buffer and applied to the reagent columns, and then washed twice with reagent buffer containing ethanol. Total RNA was eluted with RNase-free water. These bone marrow RNA samples were stored at -80 °C until use. cDNA was generated from 1 μ g total RNA using Moloney murine leukemia virus reverse transcriptase (SuperScript II, Life Technologies, Carlsbad, CA, United States), plus 20 pmol/L each dNTP and 10 pmol/L oligo dT primers in a 20- μ L final reaction volume at 42 °C for 1 h. This was followed by heating at 99 °C for 5 min.

Real-time quantitative RT-PCR

A LightCycler PCR and detection system (Roche Diagnostics, Mannheim, Germany) was used for amplification. Online quantification real-time RT-PCR was then performed in glass capillaries according to the manufacturer's protocol. The cDNA was amplified in a 20- μ L PCR reaction mixture containing each dNTP (with dUTP instead of dTTP), 1 × PCR buffer, specific primers, and magnesium chloride.

For the detection of AFP, two adjacent oligonucleotide probes were used: the LightCycler Red 640 fluorophore, hAFP-LCR; (5'-CTTGCACACAAAAGCCCACTCCA-3') and a fluorophore labeled at the 3'-end with fluorescein, hAFP-FITC; (5'-TCGATCCCACTTTTTCCAAGTT-3') (Nihon Gene Research Laboratories, Sendai, Japan). The sense and antisense primers (kindly supplied by Dr. Hiroaki Nagano at Osaka University) used for the amplification of AFP were as follows: 5'-TGCAGCCAAAGT-GAAGAGGGAAGA-3') (hAFP-s) and 5'-CATAGC-GAGCAGCCCAAAGAAGAA-3' (hAFP-As). The RT-PCR amplification was carried out for one cycle of 95 °C for 10 min, followed by 35 cycles of 95 °C for 10 s, 62 °C for 15 s, and 72 °C for 15 s. The final cycle was followed by a 10-min extension step at 40 °C.

For the detection of VEGFR-1, two adjacent oligonucleotide probes were used: hVEGFR-1-LCR; 5'-TTCC-GTGTCCCCACTGCCAA-3' and hVEGFR-1-FITC; 5'-GGGAAGCTCACTGGCATGGC-3'. The sense and antisense primers for the amplification of VEGFR-1 were as follows: 5'-TCATGAATGTTTCCCTGCAA-3' (h VEGFR-1-S) and 5'-GGAGGTATGGTGCTTCCT-GA-3' (h VEGFR-1-As). These primers were designed using sequences described in a previous report^[14]. RT-PCR amplification was carried out for one cycle of 95 °C for 10 min, followed by 35 cycles of 95 °C for 10 s, 60 °C for 10 s, and 72 °C for 16 s. The final cycle was followed by a 10-min extension step at 40 °C.

For the detection of GAPDH as an internal control, two adjacent oligonucleotide probes were used: hGAPDH-LCR; 5'-TTCCGTGTCCCCACTGC-CAA-3' and hGAPDH-FITC; 5'-GGGAAGCTCACT-GGCATGGC-3'. The sense and antisense primers for the amplification of GAPDH were as follows: 5'-GCCTCCTGCACCACCAACTG-3' (hGAPDH-S) and 5'-CGACGCCTGCTTCACCACCTTCT-3' (hGAPDH- As). The RT-PCR amplification was carried out for one cycle of 95 $^{\circ}$ C for 10 min, followed by 35 cycles of 95 $^{\circ}$ C for 10 s, 60 $^{\circ}$ C for 10 s and 72 $^{\circ}$ C for 16 s. The final cycle was followed by a 10-min extension step at 40 $^{\circ}$ C.

Quantification analysis

Quantification data were analyzed using the LightCycler analysis software (Roche Diagnostics, Mannheim, Germany) in accordance with the manufacturer's instructions. In this analysis, the background fluorescence was removed by setting a noise band. The crossing point for the calculation of amplified PCR products was set by the intersection of the best-fit line through the log-linear lesion and the noise band. The standard curve was a plot of the "crossing point" versus the copy number of DNA fragments inserted into the cloning vector.

Statistical analysis

Cumulative survival and disease-free survival (DFS) rates were computed according to the Kaplan-Meier method and compared between groups using the Breslow-Gehan-Wilcoxon test. The Cox proportional hazards model was used for multivariate analysis. Statistical analyses using standard tests (χ^2 , *t* test) were performed where appropriate. Significance was defined as *P* < 0.05. Statistical analyses were performed using StatView 5.0 Windows (SAS Institute Inc., Cary, NY, United States).

RESULTS

Analysis of AFP mRNA levels in bone marrow

The mean AFP mRNA/GAPDH ratio in the bone marrow (BM-AFP mRNA) of HCC patients, as determined by real-time quantitative RT-PCR, was 3469.27×10^{-7} (range: 0-348 526.19 $\times 10^{-7}$). The cut-off value of the BM-AFP mRNA level was set at 1.92×10^{-7} (with reference to a previous report)^[13]. The HCC patients were then divided into two groups according to this cut-off value. Accordingly, 80 patients (70.2%) were found to be negative for BM-AFP mRNA and 34 patients (29.8%), assigned to the "High" group, were positive for this transcript.

Expression of AFP mRNA in peripheral blood

No AFP mRNA was detectable in the peripheral blood of the control patients, therefore, the cut-off value for AFP mRNA/GAPDH in the peripheral blood (PH-AFP mRNA) was set at zero. Accordingly, six patients (5.4%) were found to be positive and 105 (94.6%) were negative for AFP mRNA. Due to some sampling loss, peripheral blood samples were unavailable for three patients.

Expression of VEGFR-1 mRNA in bone marrow

The mean VEGFR-1 mRNA/GAPDH in the bone marrow (BM-VEGFR-1 mRNA) of normal controls, again determined by real-time quantitative RT-PCR measurements, was 0.1497×10^{-3} (range: 0.0212×10^{-3} to 0.3213×10^{-3}). The mean BM-VEGFR-1 mRNA level in the HCC patients was 3.8474×10^{-3} (range: 0.3481×10^{-3} to





Figure 1 Expression of vascular endothelial growth factor receptor-1 mRNA in bone marrow detected by real-time quantitative reverse transcription polymerase chain reaction. The mean VEGFR-1/GAPDH mRNA ratio in the bone marrow (BM-VEGFR-1 mRNA) of normal controls was 0.1497×10^3 (range: 0.0212×10^3 - 0.3213×10^3). The mean BM-VEGFR-1 mRNA level in HCC patients was measured at 3.8474×10^3 (range: 0.3481×10^3 - 29.5885×10^3). HCC: Hepatocellular carcinoma; VEGFR: Vascular endothelial growth factor receptor; BM: Bone marrow; GAPDH: Glyceraldehyde-3-phosphate dehydrogenase.



Figure 2 Expression of vascular endothelial growth factor receptor-1 mRNA in peripheral blood detected by real-time quantitative real-time quantitative reverse transcription polymerase chain reaction. The mean VEGFR-1/GAPDH mRNA ratio in the peripheral blood (PH-VEGFR-1 mRNA) of normal controls was 2.4944×10^3 (range: 1.0730×10^3 - 4.6958×10^3). The mean PH-VEGFR-1 mRNA level in HCC patients was 9.1285×10^3 (range: 1.2774×10^3 - 174.4928×10^3). HCC: Hepatocellular carcinoma; VEGFR: Vascular endothelial growth factor receptor; PH: Peripheral blood; GAPDH: Glyceraldehyde-3-phosphate dehydrogenase.

29.5885 × 10⁻³). The mean BM-VEGFR-1 mRNA level of all HCC patients was higher than that of the normal controls (Figure 1). The HCC patients were then divided into two groups ("High" and "Low") according to their BM-VEGFR-1 mRNA level; the cut-off value was 1.5664 × 10⁻³, which was the 25th percentile value of the BM-VEGFR-1 mRNA levels in the HCC cohort. The number of patients in the High group was 81 (71.1%) and 33 (28.9%) were assigned to the Low group.

Expression of VEGFR-1 mRNA in peripheral blood

The mean VEGFR-1 mRNA/GAPDH ratio in the peripheral blood (PH-VEGFR-1 mRNA) of the normal controls was 2.4944×10^{-3} (range: 1.0730×10^{-3} to 4.6958×10^{-3}). The mean PH-VEGFR-1 mRNA level in the HCC patients was 9.1285×10^{-3} (range: 1.2774×10^{-3} to 174.4928×10^{-3}). The PH-VEGFR-1 mRNA level of almost all HCC patients was higher than that of the normal controls (Figure 2). The HCC patients were divided into high and low groups according to their PH-VEGFR-1 mRNA level. The cut-off value was 4.0238×10^{-3} , which was in the 25th percentile of the PH-VEGFR-1 mRNA level of HCC patients. The number of patients in the high group was 78 (75.0%) with 26 (25.0%) placed in the Low group. Peripheral blood samples were available for 104 patients only.

Clinical significance of the BM- and PH-VEGFR-1, and BM- and PH-AFP mRNA levels

The status of the BM-AFP mRNA levels was correlated with microscopically detectable portal invasion, whereas that of PH-AFP mRNA was found to correlate with the serum AFP and AFP-L3 levels, the number of tumors, microscopic portal invasion, and microscopic intrahepatic metastasis (Table 1). The number of tumors, serum albumin level, and a noncancerous liver were significantly correlated with the BM-VEGFR-1 mRNA level (Table 2).

Patient outcomes

Mortality: By the end of our study, 42 of the HCC patients under analysis had died; 35 from HCC, three from liver failure and four from another malignant disease. The 1-, 2- and 3-year patient survival rates for this cohort were determined to be 92.1%, 85.9% and 78.7%, respectively.

HCC recurrence: HCC recurred in 80 patients (70.2%); in the remnant liver in 48 cases (60%), in the remnant liver and remote tissue in 20 (25%), and in the remote tissue alone in 12 (15%). The 1-, 2- and 3-year DFS rates were 67.5%, 49.8% and 34.4%, respectively. We found a significant tendency for patients who were positive for BM-AFP mRNA to experience recurrence within 1 year of their surgery compared with patients who were negative for this transcript (Table 3).

HCC classification according to VEGFR-1 and AFP mRNA status

Patients were classified into four groups according to the level/status of their BM-VEGFR-1 and BM-AFP mRNA as follows: group A (n = 23), BM-VEGFR-1/BM-AFP mRNA = low/negative; group B (n = 57) high/negative; group C (n = 10) low/positive; group D (n = 24), high/ positive. This classification was correlated with disease recurrence within or more than 1 year after surgery. Significantly, in the groups in which patients were negative for BM-AFP mRNA, only three patients (13.0%) experienced recurrence within 1 year of surgery (Table 3). Classification of the HCC cases in the current study cohort by their PH-VEGFR-1 and BM-AFP (P = 0.1024), BM-VEGFR-1 and PH-AFP (P = 0.2100), and

Table 1 Characteristics of hepatocellular carcinoma patients according to their bone marrow- α -fetoprotein and peripheral blood- α -fetoprotein mRNA profile

		BM-AFP mRNA		P value	PH-AI	P mRNA	<i>P</i> value	
		Positive $(n = 34)$	Negative $(n = 80)$	-	Positive $(n = 6)$	Negative $(n = 105)$		
Sex	Male	27	67	0.5774	5	86	0.9294	
	Female	7	13		1	19		
Age (vr)	≤ 60	19	34	0.1900	5	47	0.0655	
0 0 /	> 60	15	46		1	58		
HBsAg	+	19	30	0.0697	3	45	0.7312	
0	-	15	50		3	60		
HCV	+	12	34	0.4731	2	43	0.7116	
	-	22	46		4	62		
Albumin	$\leq 4.0 \text{ mg/dL}$	13	33	0.7641	3	41	0.5937	
	> 4.0 mg/dL	21	47		3	64		
Total bilirubin	$\leq 0.7 \text{mg/dL}$	21	45	0.5853	4	60	0.6461	
	$\geq 0.8 \text{ mg/dL}$	13	35		2	45		
ICGR15	≤ 15%	22	39	0.1181	3	56	0.8736	
	> 15%	12	41		3	49		
Anatomical resection	Yes	25	54	0.5231	4	73	0.8826	
	No	9	26		2	32		
AFP	$\leq 200 \text{ ng/mL}$	21	57	0.3189	1	76	0.0040	
	> 200 ng/nL	13	23		5	29		
AFPL3	≤ 15%	21	58	0.2556	2	76	0.0418	
	> 15%	13	22		4	29		
PIVKA- II	$\leq 40 mAU/mL$	8	30	0.1477	1	36	0.3732	
	>40 mAU/mL	26	50		5	69		
Tumor number	Solitary	25	57	0.8804	2	78	0.0259	
	Multiple	9	22		4	26		
Tumor size	$\leq 2 \text{ cm}$	2	10	0.2922	0	12	0.3806	
	> 2 cm	32	70		6	93		
Differentiation	Well	0	7	0.0737	0	7	0.5859	
	Moderately	26	49		3	70		
	Poorly	7	24		3	27		
vp	Positive	14	18	0.0423	5	25	0.0014	
	Negative	20	62		1	80		
VV	Positive	3	4	0.4366	1	5	0.2098	
	Negative	31	76		5	100		
im	Positive	12	18	0.1558	4	25	0.0201	
	Negative	22	62		2	80		
Noncancerous liver		11	28	0.6833	1	36	0.3501	
Liver cirrhosis								
Non liver cirrhosis		23	49		5	66		

BM: Bone marrow; PH: Peripheral blood; AFP: α-fetoprotein; HBsAg: Hepatitis B surface antigen; HCV: Anti-hepatitis C virus antibody; AR: Patients who underwent anatomical resection; ICGR15: Indocyanine green retention rate at 15 min; vp: Microscopic tumor thrombus in the portal vein; vv: Microscopic tumor thrombus in the hepatic vein; im: Microscopic intrahepatic metastasis.

PH-VEGFR-1 and PH-AFP (P = 0.2138) mRNA status showed no such correlation. The DFS curve of group A was significantly better than that of group B, C or D (P = 0.0437, P = 0.0325, P = 0.0225, respectively; Figure 3).

Univariate analysis further revealed that age, hepatitis B surface antigen (HBsAg), albumin, AFP, AFPL3, PIV-KA-II, the number of tumors, tumor size, portal vein invasion, hepatic vein invasion, intrahepatic metastasis, BM-AFP mRNA and classification by BM-VEGFR-1/ BM-AFP mRNA are important risk factors for HCC early recurrence (Table 3). Multivariate analysis revealed that albumin ≤ 4.0 mg/dL and positive portal vein invasion were independent risk factors for recurrence within 1 year of surgery. Although BM-AFP mRNA positivity was not a significant factor by multivariate analysis, it was still found to be an important factor in predicting an early recurrence in HCC cases (P = 0.0761, Table 4).

DISCUSSION

In our current study, we found a significant tendency for HCC patients who were positive for BM-AFP mRNA to experience disease recurrence within 1 year of surgery. Patients with low BM-VEGFR-1 mRNA and who were negative for BM-AFP mRNA experienced early recurrence in 3/23 cases, whereas in 57 cases with high BM-VEGFR-1 and BM-AFP mRNA, 17 recurrences were observed. Hence, BM-AFP mRNA positivity is an important predictor of early HCC recurrence after curative hepatectomy due to hematogenic spread. BM-VEGFR-1 mRNA was also found to be associated with early HCC recurrence.

The time between hepatectomy and recurrence of metachronous *de novo* tumors is longer than that of intrahepatic metastases^[15], therefore, early recurrence of these

Table 2 Characteristics of hepatocellular carcinoma patients according to their bone marrow-vascular endothelial growth factor receptor-1 and peripheral blood-vascular endothelial growth factor receptor-1 mRNA profile

ImplementationImplementationImplementationImplementationImplementationImplementationImplementationImplementationSexMale67270.90064220.7647Age (yr) ≤ 60 40130.332237120.9097 ≥ 60 41204114141414HSAg $=$ 4619441414HCV $=$ 46100.16283290.5624Albumin $\leq 4.0 mg/dt$ 3880.025227120.2966 $\geq 4.0 mg/dt$ 4325140.37831110.378Total bilirubin $\approx 0.7 mg/dt$ 48180.613951110.378 $\geq 0.8 mg/dt$ 33157151515ICGR15 $< 15\%$ 41200.332244130.5695 $\geq 1.5\%$ 41200.332244130.5695 $\geq 0.8 mg/dt$ 33157160.56380.565ICGR15 $< 15\%$ 41200.3322840.526816 $\sim 0.8 mg/dt$ 2692280.56380.563160.5638 $\sim 100 mg/mt$ 27921100.56380.563160.5638 $\sim 100 mg/mt$ 53240.527857160.56380.563160.5638 $\sim 100 mg/mt$ 26 <th></th> <th></th> <th colspan="2">BM-VEGFR1</th> <th><i>P</i> value</th> <th colspan="2">PH-VEGFR1</th> <th><i>P</i> value</th>			BM-VEGFR1		<i>P</i> value	PH-VEGFR1		<i>P</i> value
$\begin{array}{c c c c c c c c c c c c c c c c c c c $			High $(n = 81)$	Low $(n = 33)$		High $(n = 78)$	Low $(n = 26)$	
Age (yr)Female146144Age (yr)< 60	Sex	Male	67	27	0.9090	64	22	0.7647
Age (yr) ≤ 60 40130.332237120.9097> >604120411111111111HBsAg+35140.938734120.8197HCV-46194414120.8197HCV+36100.16283290.5624-45236171418Alburnin $\leq 0.0 mg/dL$ 3880.63951110.0378> >40 mg/dL3315715160.639Total bilirubin $\leq 0.7 mg/dL$ 48180.643951110.0378> >0.8 mg/dL33157160.8026> >15%41200.332244130.6695> >15%41200.332251110.6051> >10 mg/nL55240.212455160.8026> >200 ng/nL54250.339959150.0802> >200 ng/nL54250.339959150.0802> >200 ng/nL53290.068551614> >200 ng/nL53290.06855160.8474> >200 ng/nL53290.06855160.8474> >200 ng/nL53290.068551614> >20 ng/nL53290.06855 <td></td> <td>Female</td> <td>14</td> <td>6</td> <td></td> <td>14</td> <td>4</td> <td></td>		Female	14	6		14	4	
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	Age (vr)	≤ 60	40	13	0.3322	37	12	0.9097
HBsAg + 35 14 0.9387 34 12 0.8197 HCV + 46 19 44 14 14 HCV + 36 10 0.1628 32 9 0.5624 Alburnin ≤ 4.0 mg/dL 38 8 0.0522 27 12 0.2926 Alburnin ≤ 0.7 mg/dL 48 18 0.6439 51 11 0.0378 CGR15 ≤ 15% 41 20 0.3322 44 13 0.626 Antomical resection Yes 55 24 0.6124 56 18 0.2657 AFP ≤ 200 ng/nL 54 24 0.5278 57 16 0.2653 AFP ≤ 15% 54 24 0.5278 57 16 0.2653 AFP ≤ 15% 54 25 0.3399 59 15 0.682 Inmor number Solitary 53 29 0.00	0.07	> 60	41	20		41	14	
nii<iiiiiiiii<ii<i<iiiiiiiiiiii<i<i<i<i<i<i<i<i<i<i<i<i<i<i<i<i<i<i<i<i<i<i<i<i<i<i<i<i<i<i<i< </td <td>HBsAg</td> <td>+</td> <td>35</td> <td>14</td> <td>0.9387</td> <td>34</td> <td>12</td> <td>0.8197</td>	HBsAg	+	35	14	0.9387	34	12	0.8197
HCV+36100.16283290.5624Albumin< 4.0 mg/dL	0	-	46	19		44	14	
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	HCV	+	36	10	0.1628	32	9	0.5624
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$		-	45	23		46	17	
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	Albumin	$\leq 4.0 \text{ mg/dL}$	38	8	0.0252	27	12	0.2926
$\begin{array}{c c c c c c c c c c c c c c c c c c c $		> 4.0 mg/dL	43	25		51	14	
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	Total bilirubin	$\leq 0.7 \text{mg/dL}$	48	18	0.6439	51	11	0.0378
$\begin{array}{cccccccccccccccccccccccccccccccccccc$		$\geq 0.8 \text{ mg/dL}$	33	15		27	15	
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	ICGR15	≤ 15%	41	20	0.3322	44	13	0.5695
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $		> 15%	40	13		34	13	
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	Anatomical resection	Yes	55	24	0.6124	56	18	0.8026
$\begin{array}{c c c c c c c c c c c c c c c c c c c $		No	26	9		22	8	
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	AFP	$\leq 200 \text{ ng/mL}$	54	24	0.5278	57	16	0.2653
$\begin{array}{c c c c c c c c c c c c c c c c c c c $		> 200 ng/nL	27	9		21	10	
$\begin{array}{c c c c c c c c } & 15\% & 27 & 8 & 19 & 11 \\ \hline PIVKA-II & \leq 40 \ mAU/mL & 24 & 14 & 0.1888 & 25 & 10 & 0.5491 \\ & > 40 \ mAU/mL & 57 & 19 & 53 & 29 & 0.0068 & 55 & 18 & 0.8867 \\ \hline Tumor number & Solitary & 53 & 29 & 0.0068 & 55 & 18 & 0.8867 \\ & Multiple & 28 & 3 & 23 & 7 & & & & & & & & & & & & & & & & & $	AFPL3	≤ 15%	54	25	0.3399	59	15	0.0802
$\begin{array}{c c c c c c c c c c c c c c c c c c c $		> 15%	27	8		19	11	
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	PIVKA- II	$\leq 40 \text{ mAU/mL}$	24	14	0.1888	25	10	0.5491
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$		> 40 mAU/mL	57	19		53	16	
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	Tumor number	Solitary	53	29	0.0068	55	18	0.8867
$\begin{array}{c c c c c c c c c c c c c c c c c c c $		Multiple	28	3		23	7	
$\begin{array}{c c c c c c c } & > 2 {\rm cm} & 72 & 30 & 68 & 24 \\ \hline \mbox{Differentiation} & Well & 4 & 3 & 0.1151 & 6 & 1 & 0.1614 \\ \hline \mbox{Moderately} & 51 & 25 & 56 & 11 \\ \hline \mbox{Poorly} & 26 & 5 & 16 & 14 \\ \hline \mbox{Positive} & 26 & 6 & 0.1337 & 21 & 7 & \\ \hline \mbox{Negative} & 55 & 27 & 57 & 19 \\ \hline \mbox{Vv} & \mbox{Positive} & 6 & 1 & 0.3773 & 4 & 1 & 0.7913 \\ \hline \mbox{Negative} & 75 & 32 & 74 & 25 \\ \hline \mbox{im} & \mbox{Positive} & 58 & 26 & 54 & 21 \\ \hline \mbox{Noncancerous liver cirrhosis} & 54 & 57 & 49 & 17 \\ \hline \end{tabular}$	Tumor size	≤ 2 cm	9	3	0.7499	10	2	0.4784
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$		> 2 cm	72	30		68	24	
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $	Differentiation	Well	4	3	0.1151	6	1	0.1614
$\begin{array}{c c c c c c c c c c c c c c c c c c c $		Moderately	51	25		56	11	
$\begin{array}{c c c c c c c c c c c c c c c c c c c $		Poorly	26	5		16	14	
Negative 55 27 57 19 vv Positive 6 1 0.3773 4 1 0.7913 Negative 75 32 74 25 im Positive 23 7 0.4296 24 5 0.2559 Noncancerous liver cirrhosis 58 26 54 21 Non liver cirrhosis 45 27 49 17	vp	Positive	26	6	0.1337	21	7	
vv Positive Negative 6 1 0.3773 4 1 0.7913 im Negative 75 32 74 25 im Positive 23 7 0.4296 24 5 0.2559 Negative 58 26 54 21 Noncancerous liver cirrhosis 34 5 0.0061 26 9 0.9962 Non liver cirrhosis 45 27 49 17	1	Negative	55	27		57	19	
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	vv	Positive	6	1	0.3773	4	1	0.7913
im Positive Negative 23 7 0.4296 24 5 0.2559 Negative 58 26 54 21 Noncancerous liver cirrhosis 34 5 0.0061 26 9 0.9962 Non liver cirrhosis 45 27 49 17		Negative	75	32		74	25	
Negative 58 26 54 21 Noncancerous liver cirrhosis 34 5 0.0061 26 9 0.9962 Non liver cirrhosis 45 27 49 17	im	Positive	23	7	0.4296	24	5	0.2559
Noncancerous liver cirrhosis 34 5 0.0061 26 9 0.9962 Non liver cirrhosis 45 27 49 17		Negative	58	26		54	21	
Non liver cirrhosis 45 27 49 17	Noncancerous liver cirrhosis		34	5	0.0061	26	9	0.9962
	Non liver cirrhosis		45	27		49	17	

BM: Bone marrow; PH: Peripheral blood; AFP: α -fetoprotein; HBsAg: Hepatitis B surface antigen; VEGFR: Vascular endothelial growth factor receptor; HCV: Anti-hepatitis C virus antibody; AR: Patients who underwent anatomical resection; ICGR15: Indocyanine green retention rate at 15 min; vp: Microscopic tumor thrombus in the portal vein; vv: Microscopic tumor thrombus in the hepatic vein; im: Microscopic intrahepatic metastasis.

lesions (within 1 year) is thought to be dependent on hematogenic spread. By real-time quantitative RT-PCR, we found in our current analyses that, although the AFP/ GAPDH mRNA ratios in the liver tissues were generally constant among normal control subjects, they were markedly different among HCC patients. This indicated highly variable AFP synthesis activity among individual HCC cells. It has been shown that high AFP mRNA levels reflect the presence of HCC cells^[13]. In our present study, the 1-year survival and DFS rates of HCC patients who were positive for AFP mRNA were 86.5% and 54.5%, respectively. Hence, we analyzed the relationship between early recurrence and the preoperative status of the BMand PH-AFP, and the BM- and PH-VEGFR-1 mRNA.

Although we found in our present experiments that the BM-AFP mRNA status significantly correlates with early HCC recurrence, the BM-VEGFR-1, PH-VEG-FR-1 and PH-AFP mRNA levels did not correlate with this outcome. However, classifying the HCC cases in our cohort using the BM-VEGFR-1/BM-AFP mRNA levels showed a correlation with early recurrence (P = 0.0228). Based on these findings, we speculate that the preoperative presence of cancer cells in the bone marrow is an important and essential driver of early HCC recurrence due to hematogenic spread, although we did not detect any changes in the AFP or VEGFR1 mRNA levels in the bone marrow and peripheral blood after surgical intervention in recurrent cases. The importance of the coexistence of disseminated cancer cells and VEGFR-1-positive hematopoietic bone marrow progenitor cells was further supported by the improved DFS curve of HCC patients that were negative for BM-AFP mRNA, and that showed low BM-VEGFR-1 transcript levels as compared with the other three patient groups. On the other hand, we surmised that the relationship between BM-VEGFR-1 mRNA and hematogenic spread in HCC

		Recurrence over 1 year (77)	Recurrence within 1 year (37)	<i>P</i> value
Sex	Male	64	30	0.7980
	Female	13	7	
Age (vr)	≤ 60	30	23	0.0200
007	> 60	47	14	
HBsAg	+	24	25	0.0002
0	-	53	12	
HCV	+	34	12	0.2322
	-	43	25	
Albumin	$\leq 4.0 \text{ mg/dL}$	24	22	0.0039
	> 4.0 mg/dL	53	15	
Total bilirubin	$\leq 0.7 \text{ mg/dL}$	47	19	0.3266
	$\geq 0.8 \text{ mg/dL}$	30	18	
ICGR15	≤ 15%	43	18	0.4708
	> 15%	34	19	
Anatomical resec-	Yes	55	24	0.4769
tion				
	No	22	13	
AFP	$\leq 200 \text{ ng/mL}$	59	19	0.0066
	> 200 ng/nL	18	18	
AFPL3	≤ 15%	58	21	0.0442
	> 15%	19	16	
PIVKA-II	$\leq 40 \text{ mAU/mL}$	32	6	0.0072
	> 40 mAU/mL	45	31	
Tumor number	Solitary	62	20	0.0021
	Multiple	14	17	
Tumor size	≤ 2 cm	36	6	0.0016
	> 2 cm	41	31	
Differentiation	Well	7	0	0.1631
	Moderately	52	13	
	Poorly	18	24	
vp	Positive	11	16	< 0.0001
-	Negative	66	21	
vv	Positive	2	5	0.0230
	Negative	75	32	
im	Positive	13	17	0.0010
	Negative	64	20	
BM VEGFR1	Low	24	9	0.4506
	High	53	28	
PH VEGFR1	Low	18	8	
	High	54	24	
BM AFP mRNA	Positive	17	17	0.0091
	Negative	60	20	
PH AFP mRNA	Positive	2	4	0.0569
	Negative	74	31	
BM-AFP mRNA/ BM-VEGFR1	Negative/low	20	3	0.0228
	Negative/high	40	17	
	Positive/low	4	6	
	Positive/high	13	11	

 Table 3 Clinical factors related to early hepatocellular carcinoma recurrence after curative hepatectomy

BM: Bone marrow; PH: Peripheral blood; AFP: α-fetoprotein; HBsAg: Hepatitis B surface antigen; VEGFR: Vascular endothelial growth factor receptor; HCV: Anti-hepatitis C virus antibody; ICGR15: Indocyanine green retention rate at 15 min; vp: Microscopic tumor thrombus in the portal vein; vv: Microscopic tumor thrombus in the hepatic vein; im: Microscopic intrahepatic metastasis.

during hematogenic recurrence was not stronger than that in gastric cancer, because it has been reported in a clinically relevant and widely used preclinical study model that blockade of VEGFR-1 activity does not affect the formation of spontaneous metastases^[16].

In our present study, the BM-VEGFR-1 mRNA level



Figure 3 Patients were classified into four groups according to the level/ status of their bone marrow-vascular endothelial growth factor receptor-1 and bone marrow- α -fetoprotein mRNA as follows: group A (n = 23), bone marrow-vascular endothelial growth factor receptor-1/bone marrow- α fetoprotein mRNA = low/negative; group B (n = 57) high/negative; group C (n = 10) low/positive; group D (n = 24), high/positive. The disease-free survival (DFS) curve of group A was significantly better than that of groups B, C or D (P = 0.0437, P = 0.0325, P = 0.0225).

of all HCC patients was higher than that in the normal controls, and the PH-VEGFR-1 mRNA levels of almost all of these patients were also higher than in the normal controls. Direct evidence for the role of the chemokine stromal-cell derived factor-1 [SDF-1, also known as chemokine CXC ligand (CXCL)12] in regulating the mobilization of proangiogenic bone marrow cells in vivo has been demonstrated by plasma elevation of SDF-1, which stimulates the mobilization of chemokine CXC receptor (CXCR) 4⁺ bone marrow cells, including hematopoietic stem cells and endothelial progenitor cells^[17,18]. SDF-1 not only promotes revascularization by engaging with CXCR4 expressed on vascular cells but also supports the mobilization of proangiogenic CXCR4⁺ VEGFR1⁺ hematopoietic cells^[19]. In contrast, Li et al^{20]} have reported a much higher expression level of the CXCL12-CXCR4 axis in HCC specimens than in adjacent, cirrhotic, adenocarcinoma or normal liver tissues. Hence, we speculate that VEGFR-1-positive hematopoietic bone marrow progenitor cells might be regulated and recruited by a mechanism similar to the SDF-1-CXCR4 pathway in most HCC patients. On the basis of our current data and the results of these earlier reports, we further predict that, in almost all patients with HCC, a pre-metastatic niche might have already been initiated by VEGFR-1-positive hematopoietic bone marrow progenitor cells. The levels of BM- and PH-VEGFR-1 mRNA were not found to correlate with early recurrence in each of the HCC patients, although BM-AFP mRNA positivity was significantly associated with early recurrence. These findings thus indicate that the initiation of a pre-metastatic niche be recognized as a first but essential step in the development of metastasis that requires the presence of disseminated cancer cells. This hypothesis is supported by our finding that patients negative for BM-AFP mRNA and with low levels of BM-VEGFR-1 mRNA show the lowest rate of recurrence among all of the groups analyzed.

 Table 4 Multivariate analyses of variables that are predictive of early hepatocellular carcinoma recurrence after curative hepatectomy

	P value	Risk ratio	95% CI
Age ≤ 60 yr	0.0899	3.147	0.836-11.838
HBsAg +	0.3601	1.821	0.504-6.571
Albumin $\leq 4.0 \text{ mg/dL}$	0.0038	6.536	1.832-23.256
AFP > 200 ng/nL	0.2571	2.330	0.539-10.067
$AFPL3 \le 15\%$	0.4379	1.869	0.385-9.090
PIVKA-II > 40 mAU/mL	0.1494	2.959	0.677-12.987
Tumor number solitary	0.9127	1.088	0.240-4.938
Tumor size > 3 cm	0.1177	3.026	0.756-12.114
vp positive	0.0069	6.639	1.681-26.219
vv positive	0.2221	0.234	0.023-2.408
im positive	0.2307	2.508	0.557-11.289
BM AFP mRNA: positive	0.0761	2.704	0.901-8.113

CI: Confidence interval; BM: Bone marrow; AFP: α -fetoprotein; HBsAg: Hepatitis B surface antigen; vp: Microscopic tumor thrombus in the portal vein; vv: Microscopic tumor thrombus in the hepatic vein; im: Microscopic intrahepatic metastasis.

It has been shown in several previous studies that the detection of micrometastases from solid tumors in bone marrow samples can be an important prognostic indica-tor with high specificity^[3-7,11]. The release of carcinoma cells from the bone marrow into the peripheral blood can be induced by cytokine treatment^[21]. Hence, the bone marrow might function as an important reservoir and a source of disseminated cancer cells that can subsequently spread into other organs. Moreover, the bone marrow itself may become altered in response to chemokines produced by the primary tumor and thereby enhance the metastatic capabilities of tumor cells that reside within it^[22]. It has been reported that VEGFR-1-positive cells promote tumor adherence and growth^[11]. VEGFR signaling is a crucial inducer of angiogenesis, enables primary tumor growth, and probably releases micrometastases from dormancy^[23]. In our current study, only the classification by BM-VEGFR-1 and BM-AFP mRNA was correlated with early recurrence. Hence, the coexistence of bone-marrow-derived hematopoietic progenitor cells that express VEGFR-1 in the bone marrow, and not in the peripheral blood, might be advantageous for various cancer cells in the bone marrow in terms of metastasis.

In conclusion, the evaluation of BM-AFP mRNA and BM-VEGFR-1 mRNA in patients with HCC shows great promise as a predictor of recurrence in curatively resected HCC.

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COMMENTS

Background

 α -fetoprotein (AFP) mRNA, which represents disseminate cancer cells, is re-

lated to recurrence of hepatocellular carcinoma (HCC). Bone-marrow-derived hematopoietic progenitor cells that express vascular endothelial growth factor receptor (VEGFR)-1 home to tumor-specific pre-metastatic sites and form cellular clusters before the arrival of tumor cells.

Research frontiers

It has been reported that simultaneous presence of isolated tumor cells and VEGFR-1 expression at pre-metastatic sites is clinically significant for disease progression in gastric cancer. With regard to HCC, there has been no study about the association between the presence of isolated cancer cells and the expression of VEGFR-1. In the present study, we tried to determine whether expression of AFP mRNA and VEGFR-1 in bone marrow and peripheral blood detected by real-time quantitative reverse transcriptase polymerase chain reaction (RT-PCR) could predict early recurrence in consecutive patients after curative hepatic resection.

Innovations and breakthroughs

There was a significant tendency for patients who were positive for AFP mRNA in bone marrow to experience recurrence within 1 year after surgery compared to those negative for AFP mRNA in bone marrow. The VEGFR-1 mRNA level in bone marrow in all HCC patients was higher than that of normal controls. It was supposed that this initiation of pre-metastatic niche represented by the high level of VEGFR-1mRNA might be recognized as only the first step and as a necessary condition for development of metastasis, and required the subsequent presence of disseminated cancer cells represented by AFP mRNA.

Applications

The evaluation of AFP mRNA and VEGFR-1 mRNA in bone marrow in patients with HCC could be very important for the prediction of recurrence of curatively resected HCC.

Peer review

This study found that the expression of AFP and VEGFR-1 mRNA in bone marrow detected by real-time quantitative RT-PCR predicted early recurrence in consecutive patients after curative hepatic resection. This finding is very important to elucidate the mechanism of metastasis and recurrence by hematogenic spread of HCC cells.

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